Efficacy and predictability of soft tissue ablation using a prototype Raman-shifted alexandrite laser

John A. Kozub
Jin-H. Shen
Karen M. Joos
Ratna Prasad
M. Shane Hutson
Efficacy and predictability of soft tissue ablation using a prototype Raman-shifted alexandrite laser

John A. Kozub,a Jin-H. Shen,b Karen M. Joos,b Ratna Prasad,b and M. Shane Hutsona,*

aVanderbilt University, Department of Physics and Astronomy, 6301 Stevenson Center, Nashville, Tennessee 37235, United States
bVanderbilt University Medical Center, Department of Ophthalmology and Vanderbilt Eye Institute, 2311 Pierce Avenue, Nashville, Tennessee 37232, United States

Abstract. Previous research showed that mid-infrared free-electron lasers could reproducibly ablate soft tissue with little collateral damage. The potential for surgical applications motivated searches for alternative tabletop lasers providing thermally confined pulses in the 6- to 7-μm wavelength range with sufficient pulse energy, stability, and reliability. Here, we evaluate a prototype Raman-shifted alexandrite laser. We measure ablation thresholds, etch rates, and collateral damage in gelatin and cornea as a function of laser wavelength (6.09, 6.27, or 6.43 μm), pulse energy (up to 3 mJ/pulse), and spot diameter (100 to 600 μm). We find modest wavelength dependence for ablation thresholds and collateral damage, with the lowest thresholds and least damage for 6.09 μm. We find a strong spot-size dependence for all metrics. When the beam is tightly focused (~100-μm diameter), ablation requires more energy, is highly variable and less efficient, and can yield large zones of mechanical damage (for pulse energies >1 mJ). When the beam is softly focused (~300-μm diameter), ablation proceeded at surgically relevant etch rates, with reasonable reproducibility (5% to 12% within a single sample), and little collateral damage. With improvements in pulse-energy stability, this prototype laser may have significant potential for soft-tissue surgical applications. © 2015 Society of Photo-Optical Instrumentation Engineers (SPIE)

Keywords: mid-infrared wavelengths; tissue ablation; amide I; amide II protein absorptions; Raman-shifted alexandrite laser.

Paper 150360PR received May 28, 2015; accepted for publication Aug. 28, 2015; published online Oct. 12, 2015.

1 Introduction

Previous studies have shown that mid-infrared (IR) free-electron lasers (FELs) can ablate soft tissues at rapid and reproducible rates.1–4 Furthermore, when the FEL wavelength was tuned to resonance with the amide I or amide II vibrational modes of protein—centered at 6.10 and 6.45 μm respectively—post-ablation histology showed just a thin zone (10 to 40 μm) of thermally damaged tissue around the crater periphery.1,2 These results spurred the push to conduct human trials of FELs for neurosurgical applications.5–7 These trials yielded favorable results, but the large size and high costs of FELs made them impractical for widespread use.

Several mid-IR laser systems have since been evaluated as alternatives. Some used nonlinear frequency conversion in solid-state crystalline materials,8,9 but the necessarily high pulse energies also led to rapid and permanent damage of the converting crystals. More recent alternatives have used stimulated Raman conversion in pure gases.10–12 Recirculation of the converting gas enables these systems to quickly and completely recover from optical breakdown, leading to more robust performance. Initial reports showed that one of these systems—a prototype Raman-shifted alexandrite (RSA) laser—was capable of ablating a variety of soft tissues at surgically relevant rates;12 however, the operating parameters of the RSA laser differ substantially from those of mid-IR FELs: a pulselength of just 25 ns12 as opposed to an FEL’s extended μs-long train of picosecond pulses;1 and a bandwidth of 15 nm full width at half maximum (FWHM) versus 200 nm.13 It is not immediately clear how these different operating parameters impact the RSA laser’s soft-tissue ablation metrics. Here, we measure the RSA laser’s key ablation metrics—threshold, etch rate, and amount of collateral damage—at several wavelengths and over a range of accessible pulse energies, spot diameters, and fluences. We chose three wavelengths in the 6- to 7-μm window because this range maximizes the partitioning of absorbed energy into tissue structural proteins.1,4 We find that there are some regions of the laser’s operational parameter space in which it ablates tissue cleanly and with acceptable reproducibility, but others in which collateral damage is extensive and crater depth is barely predictable. Within the limited range tested, the operational parameter that most strongly determines the accuracy and predictability of ablation is not wavelength, but is instead the combination of pulse energy and focused spot diameter.

2 Materials and Methods

2.1 Laser Parameters and Operation

Design details of the prototype RSA laser have been previously published.12 Summarizing its operational characteristics, the laser’s mid-IR output was continuously tunable from ~6.0 to 6.5 μm with a bandwidth of ~15 nm FWHM, a pulselength of 25 ns, and a repetition rate of 10 Hz. The laser emission spectrum was measured using a calibrated monochromator setup...
with a linear pyroelectric array detector (Spiricon LP-256-12-
BaF₂, Ophir-Spiricon LLC, Logan, Utah). The laser pulsewidth was measured using a fast photoelectromagnetic detector (PEM-
1, Boston Electronics, Brookline, Massachusetts). The final
multipass Raman converter was aligned daily to optimize the
spatial mode of its mid-IR output using an infrared camera
(Pyrocam III, Ophir-Spiricon LLC). When optimized, the maxi-
width obtained pulse energy ranged from 1 to 3 mJ and was
strongly wavelength dependent (see Sec. 3.1 below). For the
experiments here, the laser was operated at one of three selected
center wavelengths: 6.09, 6.27, or 6.43 μm.

2.2 Sample Preparation

Gelatin samples were prepared at 10% weight/weight by adding
boiling water to powdered gelatin (Knox, Kraft Foods Inc.).
Once completely dissolved, the solution was poured into Petri
dishes, and refrigerated until just before use. Whole eyes were
freshly harvested from euthanized pigs, goats, or dogs. These
eyes were stored in a refrigerator for no more than 24 h before
use. All samples were warmed to room temperature before
experiments and kept moist during experiments. For precise
positioning with respect to the ablating laser beam, samples
were horizontally mounted on a three-dimensional linear trans-
lation stage. Whole eyes were mounted onto the stage using a
hemispherical holder in which they were held in place by moist
tissues.

2.3 Experimental Setup

Mid-IR pulses from the prototype RSA laser were reflected
downward (perpendicular to the optical table) toward the hori-
zontal sample surface. The beamline before the sample consisted of
a fast shutter (VS-25 Uniblitz, Vincent Associates, Rochester,
New York), a holographic wire-grid polarizer (WP25H-C,
Thorlabs Inc., Newton, New Jersey), a BaF₂ beamsplitter, a
plano-convex BaF₂ lens, and a final 45-deg first-surface mirror.
Before each set of experiments, the laser was adjusted for maxi-
mum mid-IR output. This output was horizontally polarized and
the actual pulse energy delivered to the sample was attenuated as
needed using the wire grid polarizer. The pulse energy was
monitored and recorded during experiments by using the beam-
splitter to direct a small fraction of each laser pulse toward a
sensitive pyroelectric energy meter (Molectron J10, Coherent
–– converted via a measured

2.4 Exposure and Analysis

For ablation threshold measurements, single laser pulses with
varying energy were each directed to a fresh spot on a sample
surface; brightfield illumination was used to visually detect
material, and the energy of each pulse was recorded along with a
“0” or “1” to indicate the absence or presence of ejected
material. Threshold energies were extracted using probit
analysis, i.e., binomial regression of the yes/no responses
using an underlying probability function. This analysis was con-
ducted using the function ProbitModelFit in Mathematica
(Wolfram Inc., Champaign, Illinois) with probability function

Pr(ε) = \frac{1}{2} \left[ 1 + \text{erf} \left( \frac{\beta_0 + \beta_1 \epsilon}{\sqrt{2}} \right) \right],

where \( \epsilon \) is the pulse energy and \( \beta_0 \) and \( \beta_1 \) are the fit parameters.
Threshold was taken as the pulse energy that yielded a 50% probability of producing a visible ablation plume: \( \epsilon_{th} = -\beta_0/\beta_1 \).

For etch rate measurements, a two-dimensional grid of cra-
ters was made on each sample surface using a fixed nominal
generous. Craters were spaced at least 1 mm apart with each crater in a single row being exposed to the same number of pulses. The number of pulses used was then varied from one
to the next. During these exposures, the sample surface was
periodically moistered with normal saline, and excess moisture
was removed with a tissue. Immediately after ablation, the sam-
ple was imaged via optical coherence tomography (OCT) to
determine crater depths and variability. To evaluate collateral
damage, some exposed and OCT-imaged corneas were then
fixed for 24 h in Penfix (Richard-Allan Scientific, Kalamazoo,
Michigan), processed for paraffin embedding, cut into 6-μm
thick cross sections, and stained with hematoxylin and eosin
(H&E).

3 Results

3.1 Obtainable Laser Pulse Energies

As shown in Fig. 1(a), the tunable range of the RSA laser’s mid-
IR output (6.0 to 6.5 μm) corresponds to a spectral region con-
taining the amide I and II bands of tissue protein—centered
at 6.10 and 6.45 μm—and the overlapping water bending mode—
also centered at 6.10 μm. This is also the same region in which
atmospheric water vapor has a dense thicket of narrow ro-vibra-
tional bands. We find a negative correlation between this latter
spectrum and the RSA laser’s tuning curve. For example, when
the laser was set to a center wavelength of 6.10 μm, the maxi-
mum obtainable pulse energy was around 1.0 mJ, but a slight
detuning to 6.09 μm, which falls in a gap between ro-vibrational
lines, raised this to around 2.0 mJ. This problem becomes par-
icularly acute near 6.45 μm at the long wavelength end of the
laser’s tunable range. After exploring detuning to longer and
shorter wavelengths, we found the best performance for a center
wavelength of 6.43 μm, but even here the maximum obtainable
pulse energy was just 1.0 mJ. We often could not get any laser
output centered at 6.45 μm. For subsequent experiments, we
thus operated the laser at the selected center wavelengths of
6.09 and 6.43 μm. Given that the targeted tissue absorption
bands are very broad (each ~200 nm wide), we expect shifts of
just 10 to 20 nm from the tissue absorption peaks to have
negligible effects on tissue ablation metrics. To the pair of wave-
lengths selected on the above basis, we added 6.27 μm because...
it falls in the large gap between the P and R branches of the water vapor ro-vibrational spectrum. At this wavelength, we could use either a 100-mm or 200-mm focal-length lens that yielded beam waist diameters $w = 110 \pm 20$ or $290 \pm 20 \mu m$, respectively. We will refer to these as the 100- and 300-μm beam waists. As this table demonstrates, for a given sample and beam waist, there is significant wavelength dependence. The lowest thresholds were always obtained at the wavelength absorbed most strongly by both cornea and gelatin ($6.09 \mu m$). There is also an interesting spot-size dependence. As one would expect, the threshold energies increased with beam waist diameter; however, they did not increase as much as the ~9-fold increase in irradiated area. The result is that larger beam waists actually led to a decrease in the threshold fluences. For cornea, this decrease was substantial, dropping from 0.65–1.35 J/cm² to 0.16–0.62 J/cm².

To further investigate the spot-size dependence, we increased the spot diameter by displacing the sample surface away from the beam waist. As shown in Fig. 2(b), for all wavelengths and both samples, the threshold energies increased with spot size, but not as quickly as the irradiated area, again implying that threshold fluence decreased with increasing spot size. This trend is clearly evident over a wide range of spot sizes for ablation of gelatin at a wavelength of $6.09 \mu m$. We tested fewer spot sizes for other wavelengths and for cornea samples. Nonetheless, as shown in Fig. 2(c), each combination of sample and wavelength with more than one tested spot size had lower threshold fluence at larger spot diameters. In fact, the decrease in threshold for larger spots was stronger for cornea than for gelatin. We also tested the spot-size dependence for ablation of water at $6.09 \mu m$ and found that it was very weak (note the different vertical scales in Figs. 2(c) and 2(d)—varying between 0.15 and 0.3 J/cm² with only a slight decreasing trend despite a 3.6-fold change in spot diameter. The increase in spot-size dependence from water to gelatin to cornea suggests that the underlying mechanism involves the sample’s mechanical strength.

### 3.2 Ablation Thresholds

For the three selected wavelengths, the threshold energy needed for ablation of gelatin or cornea was measured using probit analysis of brightfield illumination data. An example analysis is shown in Fig. 2(a) with threshold defined as the pulse energy yielding a 50% probability for observing ejected material.

Table 1 compiles thresholds measured at the beam waist using either a 100-mm or 200-mm focal-length lens that yielded beam waist diameters $w = 110 \pm 20$ or $290 \pm 20 \mu m$, respectively. We will refer to these as the 100- and 300-μm beam waists. As this table demonstrates, for a given sample and beam waist, there is significant wavelength dependence. The lowest thresholds were always obtained at the wavelength absorbed most strongly by both cornea and gelatin ($6.09 \mu m$). There is also an interesting spot-size dependence. As one would expect, the threshold energies increased with beam waist diameter; however, they did not increase as much as the ~9-fold increase in irradiated area. The result is that larger beam waists actually led to a decrease in the threshold fluences. For cornea, this decrease was substantial, dropping from 0.65–1.35 J/cm² to 0.16–0.62 J/cm².

To further investigate the spot-size dependence, we increased the spot diameter by displacing the sample surface away from the beam waist. As shown in Fig. 2(b), for all wavelengths and both samples, the threshold energies increased with spot size, but not as quickly as the irradiated area, again implying that threshold fluence decreased with increasing spot size. This trend is clearly evident over a wide range of spot sizes for ablation of gelatin at a wavelength of $6.09 \mu m$. We tested fewer spot sizes for other wavelengths and for cornea samples. Nonetheless, as shown in Fig. 2(c), each combination of sample and wavelength with more than one tested spot size had lower threshold fluence at larger spot diameters. In fact, the decrease in threshold for larger spots was stronger for cornea than for gelatin. We also tested the spot-size dependence for ablation of water at $6.09 \mu m$ and found that it was very weak (note the different vertical scales in Figs. 2(c) and 2(d)—varying between 0.15 and 0.3 J/cm² with only a slight decreasing trend despite a 3.6-fold change in spot diameter. The increase in spot-size dependence from water to gelatin to cornea suggests that the underlying mechanism involves the sample’s mechanical strength.

### 3.3 Etch Rates and Variability with Soft Focusing

The spot-size dependence becomes even stronger for measurements of etch rates and their variability. We thus present the results separately for our two beam waist diameters—first for “soft-focusing” conditions using a 200-mm focal length lens and placing the sample surface at the beam’s 300-μm waist. Under these conditions, the focused beam had a calculated Rayleigh range of 10.8 ± 1.5 mm and its calculated diameter remained within 10% of its waist size for 5.0 ± 0.7 mm. These calculated values are consistent with measurements of the beam diameter made before and after the beam waist. Note that both values are larger than the deepest craters evaluated.

For each gelatin or cornea sample, multiple ablation craters were made at a single wavelength and constant nominal pulse energy. These craters were made in a grid pattern with each crater in a given row made using the same number of pulses. The number of pulses then varied from one row of craters to the next. Each sample was imaged via OCT immediately after completion of its grid of ablation craters. Cross-sectional OCT
images of single rows of relatively deep craters made in gelatin and cornea are shown in Figs. 3(a) and 3(e), respectively. All craters in a single image were made using the same wavelength, nominal pulse energy, and number of pulses [different parameters were used in Fig. 3(a) versus Fig. 3(e)]. At shallow depths, the craters are roughly cylindrical in both materials. At deeper depths, the craters made in gelatin maintain a nearly constant width, but those in cornea develop a markedly conical shape.

There were instances in which one or more craters penetrated through the entire cornea. When this occurred, fluid from the aqueous humor could leak out onto the corneal surface, absorb laser light delivered to nearby locations, and severely retard the ablation rate (sometimes completely preventing ablation of the cornea itself). This could lead to variations in crater depth up to 100%, but these are artifacts. Subsequent analysis thus excluded any craters that penetrated through a cornea, as well as any craters made after a perforation.

The OCT images were used to measure the depths of all partial-thickness ablation craters. When using nominally identical laser parameters, we found quite consistent crater depths within a single sample (standard deviation ~5% of the mean depth in gelatin, ~12% in cornea). This consistency held despite some targeted spots being about 750 μm past the beam waist due to curvature of the cornea surface [see Fig. 3(d)]. On the other hand, crater depths in cornea could differ by as much as a factor of 2 between samples. We did use corneas from several species (dog, pig, and goat), but the large sample-to-sample variations were not correlated with the species of origin; the variations were as large within samples from a single species as between those of different species. For subsequent analysis, we thus grouped cornea data from all three species.

For each sample, we measured multiple crater depths, Δ, made using the same wavelength, spot-size, and nominal pulse energy, but a variable number of pulses. Examples of single-sample datasets for gelatin and cornea are shown in Figs. 3(b) and 3(f), respectively. Each such dataset was fit to a simple linear regression with fixed zero offset to estimate the etch rate δ (in μm/pulse). Note that without a fixed zero offset, the large majority of linear fits would intersect the y- or Δ-axis in the region Δ > 0, indicating that crater depths increased more rapidly for the first few laser pulses before reaching a steady-state rate. Forcing the offset to zero provides a better estimate of the average etch rate as opposed to the marginal etch rate. For the two examples shown in Figs. 3(b) and 3(f), δ was 3.5 ± 0.1 and 6.1 ± 0.2 μm/pulse, as labeled on the plots. Overall for soft-focusing conditions, δ ranged from 0.5 μm/pulse to just over 10 μm/pulse, increasing with nominal pulse energy.

### Table 1: Ablation thresholds for gelatin and cornea for three Raman-shifted alexandrite laser wavelengths and two beam waist diameters.

<table>
<thead>
<tr>
<th>Wavelength (μm)</th>
<th>Gelatin thresholds (μJ)</th>
<th>Cornea thresholds (μJ)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>100-μm waist</td>
<td>300-μm waist</td>
</tr>
<tr>
<td>6.09</td>
<td>53 ± 2</td>
<td>235 ± 5</td>
</tr>
<tr>
<td>6.27</td>
<td>120 ± 5</td>
<td>—</td>
</tr>
<tr>
<td>6.43</td>
<td>96 ± 6</td>
<td>630 ± 100</td>
</tr>
</tbody>
</table>

### Fig. 2: Ablation thresholds for cornea and gelatin as a function of laser wavelength (λ) and spot diameter (2w).

(a) Example probit analysis of threshold data for cornea at λ = 6.43 μm and 2w = 300 μm. The graph shows individual measurements in which an ablation plume was either observed or not (open circles at 100% or 0 respectively), along with a probit model fit (solid line). For comparison, the measurements were also divided into eight pulse-energy bins to provide model-free estimates of the ablation probabilities (closed gray circles). Ablation threshold (50% probability of plume observation) is 0.39 ± 0.01 mJ for this example. (b) Threshold energy and (c) fluence for RSA laser ablation of gelatin (closed symbols) or cornea (open symbols) using laser wavelengths of 6.09 μm (blue circles), 6.27 μm (green squares), or 6.43 μm (red triangles). (d) Threshold fluence for RSA laser ablation of water for λ = 6.09 μm. Error bars in (b-d) denote standard errors from probit analysis and from knife-edge measurement of the laser spot size at the sample surface.
The fitted etch rates (δ in μm/pulse) and their estimated standard errors (σδ) were then compiled and compared to determine the dependence on pulse energy. Results compiled from 6 gelatin samples and 11 corneas are shown in Figs. 3(c) and 3(g). The corresponding individual crater etch rates (Δ/Np) are shown in Figs. 3(d) and 3(h) (for 285 and 450 craters respectively). As all four graphs show, there is a clear dependence on pulse energy. Unfortunately, too few independent samples were analyzed to determine whether there was a significant wavelength dependence. For the limited range of pulse energies testable—limited by ablation thresholds and by the maximum obtainable pulse energies—we used linear regression of the fitted etch rates from all wavelengths to estimate how etch rate depends on average pulse energy ⟨ε⟩. Given the large sample-to-sample variance, Figs. 3(c) and 3(g) show both the best linear fit and the 95% confidence region for predictions of etch rate in single samples. The equations corresponding to these fits for gelatin and cornea are respectively:

\[
\delta = \frac{3.5 \pm 0.1 \, \text{μm/pulse}}{\langle \epsilon \rangle - (0.3 \pm 0.4 \, \text{mJ})},
\]

The fitted etch rates (δ in μm/pulse) and their estimated standard errors (σδ) were then compiled and compared to determine the dependence on pulse energy. Results compiled from 6 gelatin samples and 11 corneas are shown in Figs. 3(c) and 3(g). The corresponding individual crater etch rates (Δ/Np) are shown in Figs. 3(d) and 3(h) (for 285 and 450 craters respectively). As all four graphs show, there is a clear dependence on pulse energy. Unfortunately, too few independent samples were analyzed to determine whether there was a significant wavelength dependence. For the limited range of pulse energies testable—limited by ablation thresholds and by the maximum obtainable pulse energies—we used linear regression of the fitted etch rates from all wavelengths to estimate how etch rate depends on average pulse energy ⟨ε⟩. Given the large sample-to-sample variance, Figs. 3(c) and 3(g) show both the best linear fit and the 95% confidence region for predictions of etch rate in single samples. The equations corresponding to these fits for gelatin and cornea are respectively:

\[
\delta = \frac{3.5 \pm 0.1 \, \text{μm/pulse}}{\langle \epsilon \rangle - (0.3 \pm 0.4 \, \text{mJ})},
\]

The fitted etch rates (δ in μm/pulse) and their estimated standard errors (σδ) were then compiled and compared to determine the dependence on pulse energy. Results compiled from 6 gelatin samples and 11 corneas are shown in Figs. 3(c) and 3(g). The corresponding individual crater etch rates (Δ/Np) are shown in Figs. 3(d) and 3(h) (for 285 and 450 craters respectively). As all four graphs show, there is a clear dependence on pulse energy. Unfortunately, too few independent samples were analyzed to determine whether there was a significant wavelength dependence. For the limited range of pulse energies testable—limited by ablation thresholds and by the maximum obtainable pulse energies—we used linear regression of the fitted etch rates from all wavelengths to estimate how etch rate depends on average pulse energy ⟨ε⟩. Given the large sample-to-sample variance, Figs. 3(c) and 3(g) show both the best linear fit and the 95% confidence region for predictions of etch rate in single samples. The equations corresponding to these fits for gelatin and cornea are respectively:

\[
\delta = \frac{3.5 \pm 0.1 \, \text{μm/pulse}}{\langle \epsilon \rangle - (0.3 \pm 0.4 \, \text{mJ})},
\]

The fitted etch rates (δ in μm/pulse) and their estimated standard errors (σδ) were then compiled and compared to determine the dependence on pulse energy. Results compiled from 6 gelatin samples and 11 corneas are shown in Figs. 3(c) and 3(g). The corresponding individual crater etch rates (Δ/Np) are shown in Figs. 3(d) and 3(h) (for 285 and 450 craters respectively). As all four graphs show, there is a clear dependence on pulse energy. Unfortunately, too few independent samples were analyzed to determine whether there was a significant wavelength dependence. For the limited range of pulse energies testable—limited by ablation thresholds and by the maximum obtainable pulse energies—we used linear regression of the fitted etch rates from all wavelengths to estimate how etch rate depends on average pulse energy ⟨ε⟩. Given the large sample-to-sample variance, Figs. 3(c) and 3(g) show both the best linear fit and the 95% confidence region for predictions of etch rate in single samples. The equations corresponding to these fits for gelatin and cornea are respectively:

\[
\delta = \frac{3.5 \pm 0.1 \, \text{μm/pulse}}{\langle \epsilon \rangle - (0.3 \pm 0.4 \, \text{mJ})},
\]
\[ \delta_c = \left( 3.4 \pm 1.6 \, \text{\textmu m/pulse/mJ} \right) \left( \langle x \rangle - (0.0 \pm 0.9 \, \text{mJ}) \right). \]

Interestingly, the slope of the pulse-energy dependence was larger for the mechanically stronger cornea samples.

### 3.4 Collateral Damage with Soft Focusing

We used histology based on H&E staining to evaluate collateral damage zones around the ablation craters. The craters evaluated for histology under soft-focusing conditions each used 50 pulses at a constant nominal pulse energy of 1.2 mJ. Representative results for our three selected wavelengths are shown in Fig. 4.

All three wavelengths are absorbed by both tissue water and protein, but with different degrees of energy partitioning. Among the three wavelengths tested, 6.43 μm yields the largest absorbed energy density in protein; \( \lambda = 6.27 \) μm yields the least. Despite these differences, we observe similar patterns of collateral damage. The example shown in Fig. 4(a) for 6.09-μm ablation displays the consistently observed structural changes: a slight swelling near the surface, a curling of the adjacent epithelial layer, and a thin darker-staining region of thermal damage along the crater margin. These features were observed for most craters at all three wavelengths with the extent of thermal damage being somewhat wavelength dependent: <10-μm thick for \( \lambda = 6.09 \) μm, 10- to 20-μm thick for \( \lambda = 6.27 \) μm [Fig. 4(b)], and nearly imperceptible for \( \lambda = 6.43 \) μm [Fig. 4(c)]. Many craters made with \( \lambda = 6.43 \) μm did, however, have a thicker (~50 μm) zone around the crater margin with a homogenized appearance [arrow in Fig. 4(c)]. Nonetheless, for all three wavelengths, the extent of thermal collateral damage was <50 μm and comparable to previous studies using FELs to ablate soft tissues. In all cases, using soft focusing, there were no discernible tears or voids indicative of mechanical damage.

### 3.5 Etch Rates and Variability with Tight Focusing

Given that the RSA laser could only produce a few mJ of energy per pulse at the tested mid-IR wavelengths, we also tested its performance when higher fluence was achieved using a more tightly focused beam. We used a 100-mm focal length lens for these tight-focusing experiments and placed the sample surface at the beam’s 100-μm waist. Under these conditions, the focused beam had a much smaller calculated Rayleigh range of 1.6 ± 0.6 mm with its diameter remaining within 10% of its waist size for just 0.7 ± 0.2 mm. These calculated values are again consistent with measurements of the beam diameter made before and after the beam waist, but are now comparable to the depth of our deepest craters.

For gelatin samples, tighter focusing yielded roughly 2× higher etch rates with variability at least comparable to, if not better than that observed above [Figs. 5(a)—5(d)]. In contrast, etch rates for cornea varied remarkably [Figs. 5(e)—5(h)]. For pulse energies <1 mJ, etch rates in cornea were usually low, just 1 to 3 μm/pulse, but already demonstrated 20% to 30% variability in crater depth. For slightly higher pulse energies, i.e., just above 1 mJ, the etch rates jumped to 10 to 20 μm/pulse. The variability in crater depths also jumped to 200% to 400%. This variability dominates the compilation of data from multiple samples and conditions shown in Figs. 5(g) and 5(h), but it is already evident in the single cornea sample shown in Fig. 5(e). Although every crater in the row shown received 20 pulses at the same wavelength (6.09 μm) and same nominal pulse energy (1.2 mJ), the crater depths varied from 150 to 750 μm.

Given the much larger etch rates for pulse energies above 1 mJ, tight-focusing conditions frequently led to rapid accidental perforation of the cornea. The data in Fig. 5(g) do include one dataset in which just 50 pulses penetrated all the way through a cornea. These data points have a single upward-pointing arrow instead of error bars to denote this measurement as a lower bound on the etch rate. As noted above, perforation of a cornea...

---

**Fig. 4** Evaluation of collateral damage from histology images (H&E staining) of ablation craters made in porcine cornea under soft-focusing conditions (2ω = 300 μm). Each crater was made at the noted wavelength using 50 laser pulses at 10 Hz and roughly 1.2 mJ per pulse. Scale bar corresponds to all three images. Arrows in the 6.43-μm panel mark the edges of a homogenized zone.
altered the sample by allowing seepage of intraocular fluid onto the corneal surface and made subsequent measurements unreliable. Subsequent analysis thus excluded craters made after a perforation; however, OCT images of suspected perforating craters often appeared to have collapsed, making it difficult to ascertain whether full perforation actually occurred. Incorrect assessments of perforation could thus contribute to the variability observed under tight-focusing conditions.

Following the same procedures for estimating the etch rate $\delta$ for single samples [Figs. 5(b) and 5(f)] and then compiling these results for all tested wavelengths again shows a clear pulse-energy dependence [Figs. 5(c) and 5(g) with 23 and 10 samples, respectively]. The same trend is evident in a compilation of all individual crater etch rates, $\Delta N_p$, for all craters. Thin horizontal lines represent craters for which individual pulse energies were not measured during exposure and extend from the average pulse energy (measured before ablation) ± one standard deviation. Thick solid lines and shaded regions are an overlay of the regression from (c, g).

Fig. 5 Crater depth measurements for gelatin (a-d) and cornea (e-h) under tight-focusing conditions—i.e., spot diameter $2w = 100 \mu m$. (a, e) Typical OCT images each showing a row of ablation craters made using the same nominal exposure conditions: $\lambda = 6.09 \mu m$ and average pulse energy $\langle e \rangle = 1.2 \text{ mJ}$. Gelatin craters were each made with 100 pulses; cornea craters were each made with just 20 pulses. Scale bars are both 1000 $\mu m$. (b, f) Example progressions of crater depth ($\Delta$) versus number of pulses delivered. Open circles show depths of individual craters; solid lines are the best linear fits to $\Delta(N_p)$ with fixed zero offset; shaded regions correspond to 95% confidence limits for single crater predictions. These examples used the same exposure conditions ($\lambda = 6.27 \mu m$ and $\langle e \rangle = 1.2 \text{ mJ}$) to yield very different fitted etch rates: $\delta = 4.70 \pm 0.03 \mu m/\text{pulse}$ in gelatin, but $10.6 \pm 0.7 \mu m/\text{pulse}$ in cornea. (c, g) Compilations from 16 gelatin samples and 10 corneas of fitted etch rates as a function of average pulse energy. Symbols denote laser wavelength as described in the legend (also applies to d, g, and h). Thick solid lines are the best linear fits to $\langle \delta(e) \rangle$ for all three wavelengths combined; shaded regions correspond to 95% confidence limits. (d, h) Compilation of individual etch rates, $\Delta N_p$, for all craters. Thin horizontal lines represent craters for which individual pulse energies were not measured during exposure and extend from the average pulse energy (measured before ablation) ± one standard deviation. Thick solid lines and shaded regions are an overlay of the regression from (c, g).
of etch rate on average pulse energy \( \langle \varepsilon \rangle \) is described by the following equations:

\[
\delta_h = \left( 3.8 \pm 0.4 \frac{\mu m/pulse}{mJ} \right) \left( \langle \varepsilon \rangle - (0.0 \pm 0.1 \ mJ) \right),
\]

\[
\delta_c = \left( 17.5 \pm 7.1 \frac{\mu m/pulse}{mJ} \right) \left( \langle \varepsilon \rangle - (0.4 \pm 0.3 \ mJ) \right).
\]

The slope of the pulse-energy dependence for gelatin increased by less than a factor of 2 under tight-focusing conditions, but it increased by a factor of 5 for cornea. Part of the variability observed among crater depths in cornea under tight-focusing conditions can thus be traced to a combination of this steep pulse-energy dependence and the laser’s 25% to 45% pulse-to-pulse energy variability. Additional sources include pulse-to-pulse changes in the beam’s spatial profile. Regardless of the exact source, it is clear that tight-focusing conditions lead to extreme variability in crater depth.

For ablation of gelatin under tight-focusing conditions, we measured etch rates for enough independent samples to evaluate the wavelength dependence (8, 8, and 7 samples, respectively, for \( \lambda = 6.09, 6.27, \) and 6.43 \( \mu m \)). To do so, we compared the wavelength independent fit above to a wavelength-dependent linear regression. The wavelength-dependent model yielded a fit with smaller residuals, but at the expense of more parameters (6 versus 2). To compare relative model quality, we used the Akaike Information Criterion, AICc, which is corrected for finite sample size.\(^{17} \) AICc was lower for the two-parameter wavelength independent model, indicating that this model led to less information loss and was thus a preferred description of the data. This result does not necessarily mean that there is no underlying wavelength dependence, only that one is not discernable given a still limited number of independent samples and large sample-to-sample variability.

### 3.6 Collateral Damage with Tight Focusing

Given the irreproducibility of cornea ablation using tight-focusing and pulse energies above 1 mJ, it is clear that such conditions are not suitable for surgical applications. We thus did not undertake an extensive analysis of collateral damage for all three selected wavelengths, but instead conducted a limited analysis just for \( \lambda = 6.09 \ \mu m \). Fig. 6 shows both OCT and histological images of craters made in a cornea using this wavelength and a constant nominal pulse energy of 1.5 mJ. The OCT image taken immediately after ablation shows increased light scattering and swelling around each crater [Fig. 6(a)]. Later histology of fixed samples showed no observable increase in H&E staining along the crater walls, but did show structural changes indicating extensive mechanical damage [Fig. 6(b)]. We observed a roughly 100-\( \mu m \) wide zone around the sides, and sometimes the bottom of the craters, in which the tissue had multiple fractures and wavy regions of altered density. These fractures were parallel to the corneal surface, suggesting delamination between stromal lamellae. The disruption in tissue structure was also associated with an increased incidence of sample tearing when making histological sections, suggesting a weakened tissue structure. Such extensive mechanical damage was not observed for tight-focusing conditions using \( \lambda = 6.09 \ \mu m \) when the pulse energy was <1 mJ.

### 4 Discussion

We have measured the characteristic ablation metrics—threshold, etch rate, and collateral damage—using a prototype mid-IR RSA laser to ablate both a mechanically weak soft tissue analog (gelatin) and a mechanically strong soft tissue (cornea). The two samples span a three-order-of-magnitude range of ultimate tensile strengths, \( \sim 10 \ \text{kPa} \) to \( 10 \ \text{MPa} \), that matches the range from highly cellularized tissues such as liver and kidney parenchyma to connective tissues like skin.\(^{20} \)

The wavelengths chosen for these experiments—6.09, 6.27, and 6.43 \( \mu m \)—all lie within a set of overlapping water and protein vibrational modes, but were slightly detuned from the wavelengths used in previous studies using mid-IR FELs. Such detuning was necessary to shift the narrow spectral output of the RSA laser to gaps between water vapor ro-vibrational lines. The specific wavelength dependence of the obtainable RSA laser performance can be attributed to a combination of a long-wavelength decline in power available from the primary alexandrite laser and absorption losses inside the system’s final multipass H\(_2\)O convertor. The latter is the stage in which near-IR light undergoes stimulated Raman conversion to the 6.0- to 6.5-\( \mu m \) range. Considering the 1.3-m length of each pass through the convertor, its nonlinear gain process can be effectively poisoned by absorption losses from just trace amounts of water vapor.\(^{21} \) Previous FEL studies did not need to explore similar detuning because the FEL linewidth was much broader, \( \sim 200 \ \text{nm} \), and thus overlapped multiple water vapor lines,
as well as the gaps between them.22 Given the ~200-nm breadth of the targeted absorption features in soft tissues, we expect the slight wavelength shifts used here to yield no significant differences in tissue ablation metrics.

In fact, overall, we find limited wavelength dependence. The ablation thresholds are certainly wavelength dependent with the lowest values obtained for \( \lambda = 6.09 \, \mu m \). This follows as expected from this wavelength’s larger absorption coefficient [Fig. 1(a)]. Our histological analysis also suggests subtle wavelength differences with \( \lambda = 6.09 \, \mu m \) leaving behind slightly less thermal collateral damage. As for the etch rates, we only had a sufficient number of samples to evaluate a wavelength dependence for gelatin under tight-focusing conditions. For this combination, an information theoretic approach showed that any wavelength dependence was indiscernible.

In contrast, we find an exceptional spot-size dependence. The ablation threshold energy increased with spot size, but at a slower rate than the irradiated area, leading to threshold fluences that decreased with spot size. Considered from another perspective, this means that more tightly focused beams required a higher fluence to initiate tissue removal. Similarly, tighter focusing for a given pulse energy increased the etch rate—by a factor of 2 to 6 depending on sample type and pulse energy—but these increases were less than the nearly 9-fold increase in fluence, meaning that ablation under tight-focusing conditions was substantially less efficient. It was also more highly variable. Finally, the degree of collateral damage observed for pulse energies >1 mJ drastically increased under tight-focusing conditions. All of these observations imply that one cannot simply compensate for limited pulse energy by focusing the beam more tightly. Beam waists \( w < 150 \, \mu m \) do not appear to be suitable for precision ablation using mid-IR lasers in surgical applications; either the etch rates are too low (for \( \varepsilon < 1 \, mJ \)) or variability and collateral damage are too high (\( \varepsilon > 1 \, mJ \)).

Previous studies of soft tissue ablation using mid-IR FELs and Er:YAG lasers demonstrated similar spot-size dependences for etch rates and thresholds.23 These studies attributed the spot-size dependence to hydrodynamic effects involved in postpulse, recoil-induced material ejection.23-26 These studies noted no spot-size dependence for collateral damage, but only when fluence was held constant; when fluence increased, so did collateral damage.23 This is consistent with our results in which tight focusing increased the fluence by a factor of 9 and led to drastically more mechanical damage. The evidence of delamination and mechanical tearing seen in these samples is also consistent recoil-induced material ejection that is constrained by the tissue’s mechanical strength.23

Despite the fact that the spot-size dependence is generally consistent with previous observations, tight focusing leads to an extremely steep dependence on pulse energy—a dependence that is detrimental to precision surgery. On the other hand, if one limits laser operation to soft-focusing conditions, then the prototype RSA laser is capable of ablating soft tissues at surgically relevant rates and with reasonable reproducibility. The variability in crater depth within a single sample at the same nominal conditions was as low as 5% for our homogeneous gelatin sample and only increased to 12% for cornea. These values represent the current status of the prototype laser’s stability-limited performance. Much of the within-sample crater variability can be attributed to fluctuations in the total amount of energy actually delivered. With improvements to the laser’s pulse energy and spatial-profile stability, it may achieve the level of reproducibility and predictability needed for precision surgical applications.

Nonetheless, even with zero fluctuations in laser parameters, natural variability in the mechanical properties of living tissue remains a problem. This is clearly evident—even using soft focusing—in the 2-fold differences in crater depth observed using nominally identical laser parameters on different cornea samples. The ideal situation for precise surgical procedures would employ real-time measurements of crater depth, allowing interactive cutting, and computer-guided beam delivery. Such interactive techniques are feasible, and prototypes have been demonstrated.26,29 Coupling these techniques to a light source similar to the prototype RSA tested here could provide an extremely useful surgical tool.

Acknowledgments

This work supported by: Department of Defense STTR W81XWH-08-C-0109; NIH Core under Grant No. 5P30 EY08126-27; unrestricted departmental grant from Research to Prevent Blindness, Inc., New York; and the Joseph Ellis Family Research Fund. We thank Marc Klosner and Chunbai Wu of Light Age Inc. for their assistance with maintenance and optimization of the prototype RSA laser.

References


John Kozub received his PhD in physics at Vanderbilt University in Nashville, TN, in 1995. He worked as a research engineer at the Keck Free Electron Laser Center at Vanderbilt, then served as associate director of the FEL Center from 2005 to 2008. He works with investigators in the Departments of Physics, Engineering, Chemistry, and the Vanderbilt Medical Center, investigating the interactions of lasers with a wide range of materials.

Jin Hui Shen received his PhD in ultrafast optical phenomena from Shanghai Institute of Optics and Fine Mechanics, Shanghai, China, in 1991. Currently, he is a research assistant professor at the Vanderbilt Eye Institute, Vanderbilt University, Nashville, TN. His research focuses on the development and testing of laser and optical coherence tomography probes.

Karen Joos received her PhD in anatomy and her MD at the University of Iowa, Iowa City, Iowa. She is professor of ophthalmology and chief of the Glaucoma Division at the Vanderbilt Eye Institute, Vanderbilt University, Nashville, TN. Clinically she specializes in the medical and surgical management of adult and pediatric glaucoma. Her research interest includes laser and optical coherence tomography projects.

Ratna Prasad is a research assistant III at the Vanderbilt Eye Institute, Nashville, TN. She specializes in multiple laboratory techniques including histology. She received her training at Gauhati University, India, with an MS in plant science mycology and pathology, and her PhD in science pathophysiology.

M. Shane Hutson received his PhD in biophysics from the University of Virginia. He is associate professor of physics at Vanderbilt University, Nashville, TN. His research interest includes the physics of laser–tissue interactions, the development of mid-infrared surgical lasers, and the use of laser microsurgery for probing cellular and developmental mechanics.