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Abstract. Quantifying tissue biomechanical properties can assist in detection of abnormalities and monitoring disease progression and/or response to a therapy. Optical coherence elastography (OCE) has emerged as a promising technique for noninvasively characterizing tissue biomechanical properties. Several mechanical loading techniques have been proposed to induce static or transient deformations in tissues, but each has its own areas of applications and limitations. This study demonstrates the combination of Lorentz force excitation and phase-sensitive OCE at ∼1.5 million A-lines per second to quantify the elasticity of tissue by directly imaging Lorentz force-induced elastic waves. This method of tissue excitation opens the possibility of a wide range of investigations using tissue biocurrents and conductivity for biomechanical analysis. © 2016 Society of Photo-Optical Instrumentation Engineers (SPIE) [DOI: 10.1117/1.JBO.21.9.090502]

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Quantifying tissue biomechanical properties can assist in the detection of abnormalities and monitoring progression of diseases and/or response to therapy. For example, it is well-known that malignant breast cancer is stiffer than healthy surrounding tissue. Elastography is a technique to map the local mechanical properties of tissues by measuring the deformation in response to external loading. Various elastography techniques have been developed and applied clinically, such as ultrasound elastography and magnetic resonance elastography. However, these techniques require relatively large displacement amplitudes to produce a detectable signal and have relatively poor spatial resolution ranging from mm to cm.

Optical coherence tomography (OCT) is a well-established noninvasive imaging technique with micrometer scale spatial resolution. As a functional extension of OCT, optical coherence elastography (OCE) shows great promise for biomechanical characterization of tissues due to its high spatial and temporal resolutions. Typically, OCT structural images have micrometer scale spatial resolution, but analyzing the phase of the complex OCT signal enables nanometer scale displacement sensitivity, which has enabled ultra-sensitive OCE techniques.

Various excitation techniques have been proposed for OCE. For example, a surgical needle was combined with a fiber-based OCE system to detect tissue margins. Acoustic-radiation force loading and OCE were used to demonstrate the age-related changes in the stiffness of rabbit crystalline lens and cornea biomechanical change after cross linking. Magnetomotive OCE (MM-OCE) was used to investigate resonance frequencies of tissue by modulating magnetic nanoparticles embedded in tissue with an external magnetic field. Low-amplitude elastic waves have been induced by a 532-nm laser and a micro air-pulse to investigate the elasticity of the cornea under various physiological conditions. However, there is no single excitation that is suitable for all applications and the development of new stimulation techniques might extend applications of OCE.

The ability of living tissue to conduct current has aroused great interest for researchers for decades. The induction of current in tissue has been used for various treatments in clinical practice. The electrical properties of tissue such as conductivity and permittivity are associated with its physiological and pathological conditions. For example, previous studies have shown that malignant tumor tissue has significantly higher permittivity as compared to normal tissue. Therefore, conductivity and permittivity of tissues could be used as effective markers for tissue diagnostics.

Lorentz force is generated when electrical current flows through the sample within an external magnetic field. Since biological tissues are inherently conductive, this force can be potentially used to investigate their electric property and chemical composition. The Lorentz force has been previously utilized in biomedical imaging for magneto-acoustic tomography and electrical impedance tomography. It can mechanically oscillate the tissue, which, in turn, might induce elastic waves. Thus, the Lorentz force presents another method to perform dynamic elastography based on mechanical wave propagation in biological tissues, which has not yet to be combined with OCE, and could open the possibility of wide range investigations using biocurrents and conductivity of tissue for biomechanical analysis.

Conventional elastic wave imaging OCE requires synchronization between multiple M-mode acquisitions (repeated A-lines measured as a function of time at one location) and multiple excitations for each measurement, resulting in long acquisition time. Recently, we have demonstrated a noncontact phase-sensitive OCE (PhS-OCE) technique at ∼1.5 million A-lines per second that was able to accurately quantify the elasticity of agar phantoms and an in situ porcine cornea. Multiple B-scans were acquired over the measurement region during elastic wave propagation (B–M mode) in <30 ms because of the significantly faster A-line rate. Therefore, this technique required only a single excitation, whereas previous investigations, which utilized M–B mode acquisition, needed an excitation for each OCE measurement position.

In this letter, we demonstrate a method of OCE by utilizing the Lorentz force to induce an elastic wave in tissue. The elasticity of agar phantoms of various concentrations and porcine liver tissue was quantified from the group velocity of a Lorentz force-induced elastic wave, which was imaged by a PhS-OCE system at ∼1.5 million A-lines per second.
Figure 1(a) is a schematic representation of the experimental setup. The Lorentz force OCE system was composed of a home-built phase-sensitive OCT system and a magnetic field generator, which was used for the Lorentz force excitation. The OCT acquisition and Lorentz excitation were synchronized by a trigger signal generated by the computer. The OCT system was based on a 4X buffered Fourier domain mode locking (FDML) swept source laser (OptoRes GmbH) with a scan rate of ∼1.5 MHz, central wavelength of 1316 nm, scan range of 100 nm, and output power up to 160 mW. The axial resolution and phase stability of the system in air were ∼16 μm and ∼9.5 mrad, respectively. Scanning was performed by a resonant scanner over ∼7.9 mm, resulting in a frame rate of ∼7.3 kHz.

The Lorentz force excitation system consisted of two NdFeB magnets, two copper wire electrodes, an arbitrary waveform generator, and a power amplifier. The two magnets were separated by a mounting base to create a sample space of 5.0 cm × 2.5 cm × 1.5 cm. The small area of Lorentz force excitation was approximately located at the center of the sample space, which has a magnetic field strength of ∼3600 gauss measured by a Gaussmeter (Model GM-1-ST, Alphalab Inc.). The arbitrary waveform generator output a 1.5-ms square pulse, which was then amplified up to a voltage of 20 to 60 V, depending on the conductivity of the sample. The two electrodes were separated ∼4 mm in a plastic bracket and connected to the sample. This ensured that the Lorentz force was parallel to the OCT scan direction. The current flowing through the sample was measured by a digital multimeter connected in the excitation circuit.

To simulate soft tissue samples of controlled stiffness, agar phantoms of three different concentrations (1%, 1.5%, and 2% w/w) were cast by mixing agar powder, 5% salt, and distilled water. A small amount of milk was added to increase scattering. Porcine liver was collected fresh from a local butcher shop and all experiments were performed within 24 h after obtaining the liver. The Lorentz force driving signal was 20 V in the phantoms and 60 V in the porcine liver.

The phase data was converted to displacement and corrected for sample surface motion. The Young’s modulus $E$ was calculated from the elastic wave group velocity $c_g$ by the surface wave equation for simplicity:

$$E = \frac{2\rho(1 + \nu)^3}{(0.87 + 1.12\nu)^2}c_g^2,$$

where $\rho = 1070 \text{ kg/m}^3$ was the measured density of the agar phantoms, $\rho = 1045 \text{ kg/m}^3$ was the density of the porcine liver, and $\nu = 0.499$ was the Poisson’s ratio to account for the incompressible nature of the phantoms and liver. All the samples were tested with a uniaxial mechanical compression testing system (Model 5943, Instron Corp.) after OCE measurements.

Figure 1(b) shows the vertical temporal displacement profile from the surface of the sample when a 100-Hz sinusoidal voltage was used to power the Lorentz force driving current. As confirmed by the spectrum obtained by FFT plotted in Fig. 1(c), each cycle was 10 ms, which corresponded to the period of input signal.

The elastic wave velocity for each sample was calculated by linearly fitting the propagation delays to the corresponding distances. The OCE measurement was taken aside of the Lorentz excitation area. Within the excitation region no time delay was observed from the OCE because the whole excitation region experienced a similar displacement. The reference position was chosen where wave propagation delays started to be observed. Displacement profiles from the surface of a 2% agar sample at 1, 2, and 3 mm away from the reference position along the wave propagation direction are presented in Fig. 2(a). The group velocities of the elastic wave in the 1%, 1.5%, and 2% phantoms were $1.86 ± 0.24$, $3.11 ± 0.15$, and $4.65 ± 0.27 \text{ m/s}$, respectively. Figure 1(b) shows the comparison of Young’s moduli of the agar phantoms as assessed by Lorentz force OCE and as measured by uniaxial mechanical testing. The Young’s moduli of 1%, 1.5%, and 2% agar phantoms as estimated by Lorentz force OCE were $12.4 ± 3.3$, $34.3 ± 3.2$, and $76.6 ± 8.8 \text{ kPa}$, respectively. Uniaxial mechanical testing showed that the elasticities of the 1%, 1.5%, and 2% agar phantoms were $15.6 ± 1.4$, $35.4 ± 2.0$, and $72.5 ± 3.7 \text{ kPa}$, respectively.
Figure 3 demonstrates the wave propagation in a spatially heterogeneous phantom made up of 1% agar on the left and 2% agar on the right (Video 1). The elastic wave velocities were calculated as 1.81 ± 0.13 m/s and 4.74 ± 0.21 m/s on the 1% and 2% components, respectively, showing the capability of using Lorentz force OCE to detect spatial variations in stiffness.

Porcine liver was utilized to test the feasibility of Lorentz force OCE to quantify the stiffness of tissue. Figure 4(a) shows the elastic wave propagation at different time points after excitation overlaid on the OCT structural image of a porcine liver sample (Video 1). The displacement is color mapped with blue as downward and red as upward displacement to better illustrate the wave propagation, and the Lorentz force excitation location lies at the left edge of the image. The elastic wave group velocity in the porcine liver was 1.83 ± 0.05 m/s, which translated to a Young’s modulus of 11.5 ± 0.6 kPa by Eq. (1). The stiffness of the porcine liver as measured by mechanical testing was 9.6 ± 2.3 kPa. The OCE-measured elastic wave velocity corroborates with previous investigations.

The Lorentz force is a new technique for elastographic excitation. Similar to MM-OCE, Lorentz force OCE needs an external magnetic field to induce a deformation. Lorentz excitation only relies on the internal conductive properties of tissue and the bioelectric current flow, so it is possible to be utilized in air and liquid environments. One advantage of Lorentz excitation is that the excitation frequency can be customized by changing the driving signal frequency as shown in Fig. 1(b), which makes it suitable for resonant spectroscopic study. Recent advances have made it possible to deliver the Lorentz excitation remotely.

It should be noted that the OCE results shows better correlation with mechanical compression testing for agar samples compared to the porcine liver. This is primarily due to assumptions made about the sample in Eq. (1). Since the viscosity of agar is usually very small and can be ignored, the Young’s modulus obtained in agar by OCE and mechanical compression testing have a better match while the viscosity effect can be neglected. In the liver, the viscosity may affect the wave speed for OCE measurement. For compression testing, the viscosity has almost no influence on the elasticity measurement. Therefore, OCE-measured Young modulus and compression results for biological samples typically show greater variability. The application of more robust mechanical models that can provide more accurate assessment of biomechanical properties is an avenue of our future work.

Fig. 2 (a) Vertical temporal displacement profiles at the indicated distances from the reference OCE measurement position from the surface of a 2% agar sample. (b) Comparison of Young's modulus of agar phantoms as assessed by Lorentz OCE and as measured by uniaxial mechanical testing (n = 4 samples for each concentration).

Fig. 3 (a) The OCT structural image of the heterogeneous phantom. (b) Group velocity calculated from the selected windows (Video 2) [URL: http://dx.doi.org/10.1117/1.JBO.21.9.090502.1].

Fig. 4 (a) The OCT structural image of a porcine liver sample and elastic wave propagation overlay. (b) Comparison of elasticity as assessed by Lorentz force OCE and as measured by uniaxial mechanical testing (Video 3) [URL: http://dx.doi.org/10.1117/1.JBO.21.9.090502.2].
been previously investigated using OCT. The electrical field change may cause various electro-kinetic responses such as electric field-induced mechanical changes, especially in vivo. Due the relatively weak and short duration of the electric field, these changes are typically confined locally to the excitation position, but this deserves further investigation.

We have demonstrated a stimulation technique utilizing the Lorentz force to induce an elastic wave in tissue, which was imaged by a PhS-OCE system at ~1.5 million A-lines per second. The results show that Lorentz force OCE was able to accurately assess the elasticity of tissue as compared to mechanical testing.

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